



8 as a major product strongly supports the presumption that intersystem crossing of 12 to 11 occurs.¹⁹ It would thus appear that, along with rearrangement, insertion results from reaction of 11. Without further information, however, it can not be concluded that conversion into 4 and 5 is the exclusive province of singlet 11.19

Direct photolysis and singlet photosensitization of 1 and 2 occur with absorption of \sim 82–90 kcal/mol of energy. Electronically excited singlets 1-2* and/or possibly excited 15* are thus highly energetic and their conversions into 4^* and 5^* are spin allowed. Rearrangement to 5* is now extensive and the product does not undergo alteration as occurs when derived from vibrationally excited intermediates in the gas phase.^{7e} Finally, triplet photosensitization of diazo compounds and diazirines as in the present systems may give advantage over thermolysis and direct photolysis for specific synthesis.

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- 5 (8%)
- Effected at 302.5, 313, and 334 nm with a Hanovia lamp (679A) through Pyrex

(12) In diglyme containing lithium tert-butoxide, 2 photolyzes at 20 °C to 4 (51.6%), 5 (46.6%), and 7 (1.8%). (13)

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- (16) Neopentane (~1%) formed upon photosensitizations in cumene possibly arises by hydrogen abstraction by 12 and then the neopentyl radical.
- Isomerizations of 12 with spin preservation to 13 and 14 apparently are (17)higher energy demanding processes.^{2d}
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Total Synthesis of Tryptoquivaline G



A strain of the fungus Aspergillus clavatus collected from mold damaged rice produced a group of toxic, tremor inducing metabolites with novel structures. Tryptoquivaline (1) was found to be the major metabolite, and a transformation product containing a δ -lactone ring was used to determine its structure and relative configuration by X-ray crystallography.¹ Comparison of circular dichroism and 'H NMR spectra with those of nortryptoquivaline² suggested structure 2 for this companion metabolite. Tryptoquivaline G(3) is a representative of a more recently discovered group of mycotoxins produced by Asper-gillus fumigatus.³⁻⁵ lt, as well as tryptoquivaline L (17), an artefact, lacks the isobutyl side chain. The total synthesis of tryptoquivaline G (3) outlined here confirms the proposed



structure and establishes both relative and absolute configuration.

The synthesis relied heavily on a new method for the conversion of N-acyltryptophans to spirolactones and the steric course of the reaction was explored with a model compound.⁶ Oxidation of N-phthalimido-L-tryptophan with 2 equiv of trichloromethanesulfonyl chloride-dimethyl sulfoxide⁷ (CH₂Cl₂, -20 °C) gave a 65% yield of two diastereomeric lactones (4 and 5) in a ratio of 7:3. The major isomer 4, mp



275-277 °C, $[\alpha]^{25}_{\rm D}$ -133° (c 1.7, acetone), was recovered unchanged after treatment with sodium hydride or imidazole in dimethylformamide. The minor isomer **5**, mp 279-281 °C, $[\alpha]^{24}_{\rm D}$ -206° (c 1.4, acetone), under the same conditions was converted into the *enantiomer* of **4**, $[\alpha]^{25}_{\rm D}$ +132° (c 1.4, acetone), demonstrating different configurations at the spiro atom in the two diastereomers. Since the epimerization of tryptoquivaline G (**3**) to tryptoquivaline L (**17**) is accompanied by a large negative shift in optical rotation, the absolute configurations at C₃ and C₁₉ should be opposite those of the model compound **5**. The major, and thermodynamically more stable, epimer formed in the oxidative lactonization thus has the correct stereochemistry at the spiro center.

Intermediate 6 was prepared by condensation of L-tryptophan with o-nitrobenzoyl chloride to give the amide, mp



212-214 °C, esterification with phenyldiazomethane to the benzyl ester, mp 148-150 °C, and reduction with iron (HCl, ethanol, reflux). The formamide, mp 140-142 °C (HCOOH, benzene, reflux), on dehydration (TsOH, xylene, reflux) gave a quinazolinone which was hydrogenolyzed to the acid 7, mp 238-240 °C, over palladium on carbon in ethanol (overall yield from L-tryptophan was 50%). Oxidation of 7 with 2 equiv of methanesulfonic anhydride-dimethyl sulfoxide (CH₂Cl₂, -20 °C, 5 h) gave 56-66% spirolactone 8, mp 321-322 °C dec, $[\alpha]^{25}$ _D -377 ° (c 0.07, CH₃CN), in addition to <10% epimer differing in configuration at the spiro center. To confirm the stereochemistry, 8 was exposed to potassium hydride in THF-DMF (20 °C, 3 min), and the resulting enolate was protonated with 1% HCl in THF (-70 °C). Product 9 (40%), mp 315-318 °C dec, $[\alpha]^{25}_{D}$ +320° (c 0.02, CH₃CN), was the enantiomer of the minor product formed in the oxidative lactonization. Owing to the exceptional lability of 9 to base, the

synthesis was continued with 8. Derivative 10, mp 82-83 °C, was prepared from α -methylalanine and p-methoxybenzyl-8-quinolyl carbonate⁸ with the aid of triethylamine, followed by esterification with p-nitrophenol and dicyclohexylcarbodiimide. Lactone 8 was then silylated with bis(trimethylsilyl)acetamide⁹ and the crude product condensed with 10 in DMF containing tetramethylammonium chloride. Treatment of the crude imide with triethylamine gave the highly insoluble cyclol 11, mp 243-245 °C, in 81% overall yield. Transforma-



tion to the aminals 15 and 16 could be accomplished by consecutive treatment of 11 with sodium cyanoborohydride, DDQ, and trifluoroacetic acid, but the two epimers were obtained in a ratio of 1:1. In a more stereoselective sequence, the pmethoxybenzyloxycarbonyl group was first removed by treatment of a suspension of the cyclol 11 in ethyl acetate with trifluoroacetic acid in the presence of anisole (0 °C, 30 min).¹⁰ The deprotected cyclol 12, mp 268-270 °C (76%), was reduced with sodium cyanoborohydride (THF, H₂O, HCl, pH ~3, 20 °C, 1 h) to give a 4:1 mixture of dihydroquinazolinones 13, mp 248-250 °C, and 14, mp 243-245 °C. Reoxidation to the corresponding quinazolinones 15, mp 250-251 °C (acetone), $[\alpha]^{25}_{D}$ -260° (c 0.01, acetone), and 16, mp 172-174 °C (AcOEt-hexane), $[\alpha]^{25}_{D}$ -67° (c 0.12, CHCl₃), was accomplished in 89% and 80% yield, respectively, with DDQ in chloroform. ¹H NMR spectra of the two isomers were used to establish the configurations at C_2 , and the major isomer on oxidation with m-chloroperbenzoic acid² gave tryptoquivaline L (17, 85%), mp 275–277 °C dec, $[\alpha]^{25}$ –230° (c 0.02, acetone), with spectral properties in accord with those published.^{4,5} Contrathermodynamic epimerization, as in the conversion of 8 into 9, gave tryptoquivaline G identical (melting point, $[\alpha]_D$, IR, ¹H NMR and chromatographic behavior) with material isolated from A. fumigatus.¹¹ The tryptoquivalines are thus derived from D-tryptophan.

The configuration of nortryptoquivaline (2) was established by X-ray analysis¹² and reduction of this metabolite with zinc followed by hydrolysis which yielded L-alanine.^{3,5} The absolute configuration of the metabolite (2) thus agrees with that of tryptoquivaline G (3) determined by synthesis.

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General Acid Catalysis in the Hydrolysis of Benzo[a]pyrene 7,8-Diol 9,10-Epoxides

Sir:

The most studied of the carcinogenic hydrocarbons is the ubiquitous environmental contaminant benzo[a]pyrene (BP). Studies from several laboratories¹ have recently allowed the identification of the metabolite (+)-7 β ,8 α -dihydroxy- 9α , 10α -epoxy-7, 8, 9, 10-tetrahydro BP ((+)-2, Figure 1) as an ultimate carcinogen of BP in newborn mice.² Two diastereomeric 7,8-diol 9,10-epoxides of BP are metabolically possible, each capable of existing in enantiomeric forms. Although (\pm) -1 is >30-fold more hydrolytically reactive in the physiological pH range⁵ and more susceptible to attack by nucleophiles in nonaqueous solution relative to (\pm) -2,⁶ only (+)-2 is tumorigenic in newborn mice. Both 1 and 2 alkylate the phosphate backbone of nucleic acid and effect the strand scission of DNA.7 Because alkylation at phosphate may play an important role in the mutagenic and carcinogenic activities of 1 and 2, the mechanisms of reaction of 1 and 2 with hydrogen phosphate species (and other general acids) in aqueous dioxane solutions were determined and are reported in this study.8

The rates of reaction of 1 and 2 in 10% dioxane-water solutions at a given pH exhibit a marked first-order dependence on concentration of phosphate buffer (Figure 2).^{9,10} The rate data for series of serially diluted buffer solutions of constant pH were fit to

$$k_{\rm obsd} = k_{\rm HA}[{\rm HA}] + k_{\rm H} + a_{\rm H^+} + k_0 \tag{1}$$

where $k_{\rm H}+a_{\rm H}+$ and k_0 represent contributions by the acid catalyzed and spontaneous hydrolysis mechanisms, respectively,⁵ and HA refers to the dihydrogen phosphate ion (H₂PO₄⁻). Values of $k_{\rm H_2PO_4}$ - for hydrolysis of 1 and 2, obtained from plots of $k_{\rm obsd}$ vs. [H₂PO₄⁻] for solutions at a given pH, were found to be constant within experimental error over



Figure 1. Diastereomeric BP 7.8-diol 9.10-epoxides.³ The enantiomers of each diastereomer shown are those found bound to nucleic acid when BP is applied to mouse skin.⁴



Figure 2. Plots of k_{obsd} vs. $[H_2PO_4^-]$ for the hydrolysis of 1 and 2 in 10% dioxane-water solutions at 25 °C, ionic strength 0.2 (NaClO₄), pH 7.02.

Table I. Values of k_{HA} for the General Acid Catalyzed Hydrolysis of 1 and 2 in 10% Dioxane-Water^{*a*} at 25 °C^{*b*}

НА	pН	$k_{HA}(1),$ M ^{~1} s ⁻¹	k _{HA} (2), M ^{−1} s ^{~1}
НОАс	4.81	0.72 ± 0.06	1.32 ± 0.02
	5.11	0.71 ± 0.04	1.33 ± 0.03
H ₂ PO ₄ -	6.34	0.31 ± 0.01	0.48 ± 0.03
	6.70	0.28 ± 0.03	0.49 ± 0.01
	7.02	0.27 ± 0.01	0.48 ± 0.02
	7.10	0.28 ± 0.01	0.54 ± 0.02
	7.60	0.25 ± 0.01	0.49 ± 0.02
Tris-H+	7.94	0.055 ± 0.006	0.026 ± 0.001
	8.54	0.056 ± 0.005	0.020 ± 0.002
$HO(CH_2)_2NH_3^+$	8.68	0.048 ± 0.004	0.0043 ± 0.0003
	8.94	0.048 ± 0.002	0.0044 ± 0.0004
phenol	9.22	0.19 ± 0.02	0.046 ± 0.003
	9.58	0.17 ± 0.01	0.046 ± 0.002

^{*a*} Volume/volume; ionic strength, 0.2 (NaClO₄). ^{*b*} Rates were monitored by observing the absorbance change of the reaction solutions at 348 nm in the thermostated cell compartment $(25.0 \pm 0.1 \text{ °C})$ of a Gilford 2400 spectrophotometer. Rate constants were determined from least-squares plots of k_{obsd} vs. [HA] for solutions of constant pH but varied buffer concentrations.

the pH range studied (6.3-7.6, Table I). These data indicate that $H_2PO_4^-$ is the only catalytic or reactive phosphate species in the hydrolysis of 1 and 2 under these conditions.

Several kinetically indistinguishable mechanisms might account for the $H_2PO_4^-$ term in eq 1. One possibility is the specific acid-general base mechanism outlined in Scheme I (arbitrarily shown for the reaction of 1). This mechanism involves attack of hydrogen phosphate ion (HPO_4^{-2}) on the protonated epoxide 3 to yield phosphate ester 4.